

ORIGINAL ARTICLE

헬리코박터 파일로리 음성 기능성 소화불량증 환자 위와 십이지장의 호산구와 비만세포 수

민양원*, 이혁*, 안수민¹, 송경호², 박종규³, 신철민⁴, 허규찬⁵, 대한소화기기능성질환 운동학회 산하 기능성 소화불량증 연구회

성균관대학교 의과대학 삼성서울병원 내과, 병리과¹, 차의과학대학교 일산차병원 내과², 울산대학교 의과대학 강릉아산병원 내과³, 분당서울대학교병원 내과⁴, 건양대학교 의과대학 내과학교실⁵

Eosinophil and Mast Cell Counts in the Stomach and Duodenum of Patients with Functional Dyspepsia without a *Helicobacter pylori* infection

Yang Won Min*, Hyuk Lee*, Soomin Ahn¹, Kyung Ho Song², Jong Kyu Park³, Cheol Min Shin⁴, Kyu Chan Huh⁵; and Functional Dyspepsia Study Group Under the Korean Society of Neurogastroenterology and Motility

Departments of Medicine, Pathology and Translational Medicine¹, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul; Department of Internal Medicine, CHA Ilan Medical Center, CHA University, School of Medicine², Goyang; Department of Internal Medicine, Gangneung Asan Hospital, University of Ulsan College of Medicine³, Gangneung; Department of Internal Medicine, Seoul National University Bundang Hospital⁴, Seongnam; Department of Internal Medicine, Konyang University College of Medicine⁵, Daejeon, Korea

Background/Aims: Symptom-based subtyping of functional dyspepsia (FD) is used to segregate patients into groups with homogenous pathophysiological mechanisms. This study examined whether subtyping could reflect the duodenal and gastric microinflammation in FD patients.

Methods: Twenty-one FD patients without *Helicobacter pylori* infection were recruited. An endoscopic biopsy was performed in the duodenum 2nd portion, stomach antrum, and body. The eosinophil and mast cell counts per high-power field (×40) were investigated by H&E and c-kit staining, respectively. The degree of inflammatory cell infiltration, atrophy, and intestinal metaplasia was also determined by H&E staining in the stomach. The baseline characteristics and eosinophil and mast cell infiltrations were compared among the three groups (epigastric pain syndrome, postprandial distress syndrome, and overlap).

Results: According to the symptom assessment, seven subjects were classified into the epigastric pain syndrome group, 10 into the postprandial syndrome group, and four into the overlap group. The baseline variables were similar in the three groups. Eosinophil infiltration was more prominent in the duodenum than in the stomach. In contrast, mast cell infiltration was similar in the duodenum and stomach. The eosinophil counts in the duodenum were similar in the three groups. The eosinophil counts in the stomach and mast cell counts in the duodenum and stomach were also similar in the three groups.

Conclusions: Duodenal eosinophil infiltration was prominent in FD patients, but the eosinophil counts were similar regardless of the symptom-based subtypes of FD. Hence, the current symptom-based subtyping of FD does not reflect duodenal eosinophil and mast cell infiltration. (Korean J Gastroenterol 2022;80:28-33)

Key Words: Duodenum; Dyspepsia; Eosinophils; Mast cells; Upper gastrointestinal tract

Received February 28, 2022. Revised June 21, 2022. Accepted June 22, 2022.

© This is an open access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. Copyright © 2022. Korean Society of Gastroenterology.

교신저자: 허규찬, 35365, 대전시 서구 관저동로 158, 건양대학교 의과대학 소화기내과

Correspondence to: Kyu Chan Huh, Division of Gastroenterology, Department of Internal Medicine, Konyang University College of Medicine, 158 Gwanjeodong-ro, Seo-gu, Daejeon 35365, Korea. Tel: +82-42-600-9104, Fax: +82-42-600-9058; E-mail: kchuh2020@hanmail.net, ORCID: <https://orcid.org/0000-0003-3746-8419>

Financial support: This study was financially supported by a grant of the Korean Society of Neurogastroenterology and Motility for 2014.

Conflict of interest: None.

* These authors contributed equally to this work as first authors.

INTRODUCTION

Functional dyspepsia (FD) is a common disorder characterized by the presence of chronic or recurrent symptoms of upper abdominal pain or discomfort in the absence of known specific structural causes.¹ Several pathophysiological mechanisms have been suggested to underlie dyspeptic symptoms, including impaired gastric accommodation, delayed gastric emptying, impaired mucosal permeability, low-grade inflammation, gastric hypersensitivity, *Helicobacter pylori* (*H. pylori*) infection, altered response to duodenal lipids or acid, abnormal duodenojejunal motility, and central nervous system dysfunction.²

Although targeting the underlying pathophysiological mechanisms is ideal for therapeutic strategies, tests for mechanism evaluation are usually impractical, and single predominant symptoms cannot predict disease mechanisms accurately.³⁻⁶ The Rome IV criteria suggest using symptom-based subtypes, such as postprandial stress syndrome (PDS) and epigastric pain syndrome (EPS), based on the assumption that subtyping could segregate patients with FD into homogenous mechanisms groups.⁷ Many practice guidelines refer to symptom-based subtypes for choosing the first-line treatment option, such as proton pump inhibitors for EPS and prokinetics for PDS.⁸⁻¹⁰

Over the last decade, emerging data point towards the duodenal microinflammation (most notably eosinophilic duodenitis and intestinal mast cell disease) as a crucial underlying pathophysiological mechanism of FD.¹¹⁻¹⁴ As treatments targeting gastric sensorimotor function are of limited efficacy,¹⁵⁻¹⁷ duodenal pathology should be addressed from the aspect of the therapeutic target. Thus, the present study examined the differences in eosinophil and mast cell infiltration in the duodenum and stomach according to the symptom-based subtypes of FD.

SUBJECTS AND METHODS

1. Subjects

This retrospective study included patients with FD who satisfied the Rome IV criteria and underwent upper endoscopy with gastric and duodenal 2nd portion biopsy for investigating *H. pylori* infection and duodenal inflammation. The patients were excluded if they had *H. pylori* infection or had undergone

previous abdominal surgery. The diagnosis of a *H. pylori* infection was made using Giemsa staining for the endoscopic biopsy samples from the gastric antrum and body.

This study protocol was conducted in accordance with the Declaration of Helsinki and approved by the Institutional Review Board (IRB) at Samsung Medical Center, Seoul, Korea, on the 15th of April 2013 (IRB No. 2020-12-093). The requirement for informed consent was waived because routinely collected de-identified data were used.

2. Data collection

From the chart review, the demographic characteristics (age and sex), symptoms of FD, and presence of other diseases (gastroesophageal reflux disease, irritable bowel syndrome [IBS], hypertension, and diabetes) were obtained. Subjects were classified into three symptom-based subtypes of FD (EPS, PDS, and overlap).⁷ All endoscopic images were reviewed by an experienced endoscopist (Min YW), and the status of reflux esophagitis, Hill grade,¹⁸ and chronic atrophic gastritis was determined. The chronic atrophic gastritis grade was diagnosed by evaluating the atrophic border in the endoscopic images. The atrophic pattern system described by Kimura et al.¹⁹ was used.

An experienced pathologist (Ahn S), blinded to the clinical information, investigated the gastric and duodenal pathology of previously obtained unstained biopsy samples. The eosinophil and mast cell counts per high-power field (HPF, $\times 40$) were investigated by H&E and c-kit staining, respectively. The counts were determined by the total number of eosinophils and mast cells in the five non-overlapping HPFs. The degrees of inflammatory cell infiltration, atrophy, and intestinal metaplasia were also determined by H&E staining of two biopsy sites (gastric antrum and body). Atrophy was defined as a decrease in the number of appropriate glands. Inflammatory cell infiltration and metaplastic and non-metaplastic atrophy were determined using the updated Sydney system scores (i.e., 0=none, 1=mild, 2=moderate, and 3=marked).

3. Statistical analysis

Statistical analyses were conducted using GraphPad Prism 9.2.0 software (GraphPad, San Diego, CA, USA). The baseline characteristics were compared among groups using one-way analysis of variance (ANOVA) with multiple comparisons and the χ^2 test. The cell counts were compared among groups

Table 1. Baseline Characteristics of the Study Participants according to the Symptom-based Subtypes of Functional Dyspepsia

	EPS (n=7)	PDS (n=10)	Overlap (n=4)	p-value
Age (years)	62.4±6.8	51.7±17.2	63.0±11.5	0.213
Male sex	1 (14.3)	4 (40.0)	0 (0.0)	0.218
Hypertension	0 (0.0)	2 (20.0)	0 (0.0)	0.296
Diabetes	0 (0.0)	0 (0.0)	0 (0.0)	NS
Irritable bowel syndrome	1 (14.3)	1 (10.0)	0 (0.0)	0.738
Reflux esophagitis	1 (14.3)	3 (30.0)	0 (0.0)	0.402
Hill grade				0.165
1	4 (57.1)	3 (30.0)	1 (25.0)	
2	3 (42.9)	6 (60.0)	1 (25.0)	
3	0 (0.0)	1 (10.0)	2 (50.0)	

Values are presented as number (%) or mean±standard deviation.

EPS, epigastric pain syndrome; PDS, postprandial stress syndrome; NS, not significant.

Table 2. Stomach Pathologic Characteristics of the Study Participants according to Symptom-based Subtypes of Functional Dyspepsia

	EPS (n=7)	PDS (n=10)	Overlap (n=4)	p-value
Antrum inflammation				0.552
0	4 (57.1)	7 (70.0)	3 (75.0)	
1	2 (28.6)	1 (10.0)	1 (25.0)	
2	0 (0.0)	1 (10.0)	0 (0.0)	
3	1 (14.3)	1 (10.0)	0 (0.0)	
Antrum atrophy				0.868
0	5 (71.4)	7 (70.0)	3 (75.0)	
1	1 (14.3)	1 (10.0)	1 (25.0)	
2	1 (14.3)	2 (20.0)	0 (0.0)	
3	0 (0.0)	0 (0.0)	0 (0.0)	
Antrum intestinal metaplasia				0.852
0	4 (57.1)	7 (70.0)	3 (75.0)	
1	2 (28.6)	1 (10.0)	1 (25.0)	
2	0 (0.0)	1 (10.0)	0 (0.0)	
3	1 (14.3)	1 (10.0)	0 (0.0)	
Body inflammation				0.146
0	0 (0.0)	0 (0.0)	0 (0.0)	
1	7 (100.0)	7 (70.0)	4 (100.0)	
2	0 (0.0)	3 (30.0)	0 (0.0)	
3	0 (0.0)	0 (0.0)	0 (0.0)	
Body atrophy				0.738
0	6 (85.7)	9 (90.0)	4 (100)	
1	1 (14.3)	1 (10.0)	0 (0.0)	
2	0 (0.0)	0 (0.0)	0 (0.0)	
3	0 (0.0)	0 (0.0)	0 (0.0)	
Body intestinal metaplasia				0.561
0	7 (100.0)	9 (90.0)	4 (100.0)	
1	0 (0.0)	1 (10.0)	0 (0.0)	
2	0 (0.0)	0 (0.0)	0 (0.0)	
3	0 (0.0)	0 (0.0)	0 (0.0)	

Values are presented as number (%).

EPS, epigastric pain syndrome; PDS, postprandial stress syndrome.

using the Kruskal-Wallis test and one-way ANOVA with multiple comparisons. Statistical significance was defined as a two-sided p -value <0.05 .

RESULTS

1. Subject characteristics

Twenty-one subjects were included in this study. According to the symptom assessment, seven subjects were classified into the EPS group, 10 into the PDS group, and four into the

overlap group (Table 1). The baseline characteristics (age, sex, presence of hypertension, diabetes, IBS, reflux esophagitis, and Hill grade) were similar in the three groups. The stomach pathology (inflammation, atrophy, and intestinal metaplasia in the antrum and body) was similar in the three groups (Table 2).

2. Eosinophilic count

Although the eosinophilic counts were higher in the duodenum than in the stomach (Table 3, Fig. 1A-C), no significant differences were found among the three groups in terms of

Table 3. Eosinophil and Mast Cell Counts according to the Symptom-based Subtypes of Functional Dyspepsia

	EPS (n=7)	PDS (n=10)	Overlap (n=4)	p-value
Duodenum				
Eosinophils (/HPF)	12 (7-21)	11.5 (6.25-17.75)	9.5 (7.5-13)	0.758
Mast cells (/HPF)	22 (20-26)	24 (16.5-26.5)	28.5 (27.25-29.75)	0.076
Stomach antrum				
Eosinophils (/HPF)	1 (0-5)	1.5 (0.75-5.75)	1.5 (0.25-13.25)	0.856
Mast cells (/HPF)	25 (20-29)	21.5 (10.75-23.5)	24.5 (20.75-29)	0.245
Stomach body				
Eosinophils (/HPF)	2 (1-4)	2.5 (0-3.75)	0.5 (0-1.75)	0.231
Mast cells (/HPF)	16 (15-19)	17.5 (10-19.75)	25.5 (19-29.75)	0.058

Values are presented as median (interquartile range).

EPS, epigastric pain syndrome; PDS, postprandial stress syndrome; HPF, high-power field.

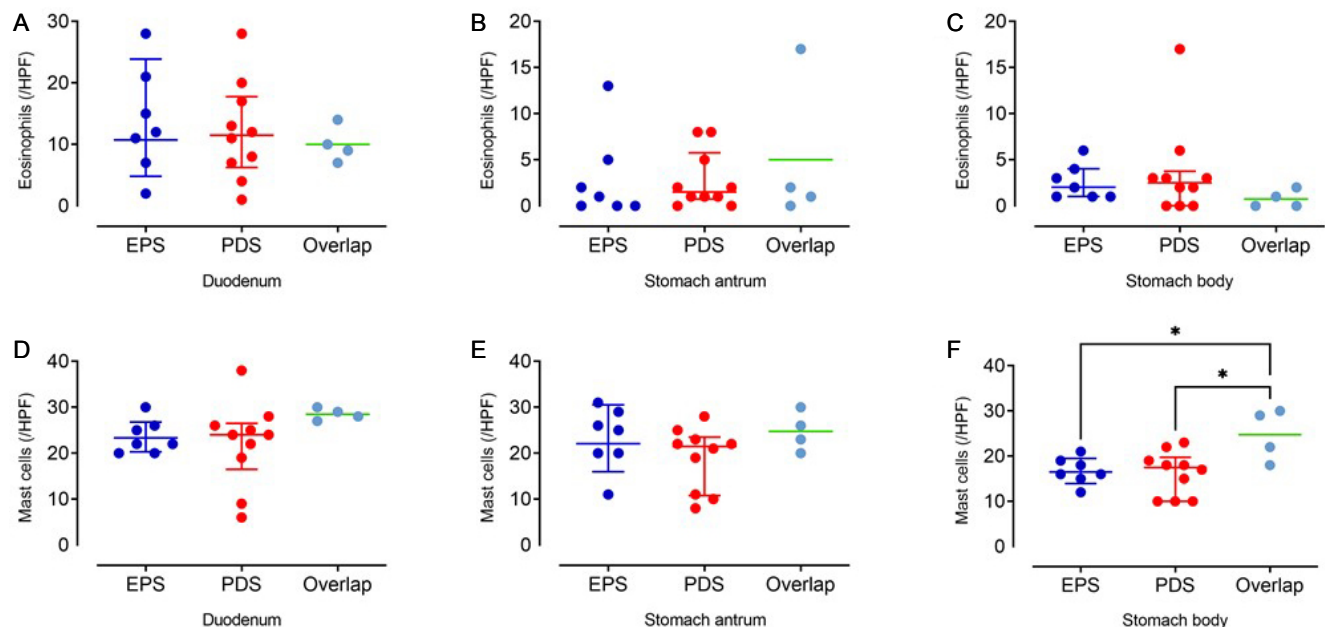


Fig. 1. Eosinophil and mast cell counts according to symptom-based subtypes of functional dyspepsia. Comparison of the eosinophil counts in the (A) duodenum, (B) stomach antrum, (C) stomach body and of mast cell counts, in the (D) duodenum, and (E) stomach antrum according to symptom-based subtypes of functional dyspepsia showed no significant difference among the groups. (F) The stomach body mast cell count was higher in the overlap group than in the EPS and PDS groups. EPS, epigastric pain syndrome; PDS, postprandial stress syndrome.

the eosinophilic counts of the duodenum ($p=0.758$) or the stomach (antrum, $p=0.856$; body, $p=0.231$).

3. Mast cell count

The mast cell counts were similar in the duodenum and the stomach (Table 3, Fig. 1D-F). Although the three groups showed similar mast cell counts in the duodenum ($p=0.076$) and stomach (antrum, $p=0.245$; body, $p=0.058$), the stomach body mast cell counts were higher in the overlap group than in the EPS and PDS groups (adjusted $p=0.027$ and $p=0.012$, respectively) (Fig. 1F).

DISCUSSION

Although FD is a common disorder worldwide, the clinical outcomes are unsatisfactory. One of the reasons might be the heterogeneous pathophysiological mechanisms in the patient group. To increase the therapeutic efficacy, physicians attempt to determine the mechanisms of the disease through symptom assessment in usual practice and perform physiological tests in selected cases. Symptom-based subtypes, such as PDS and EPS, are believed to be more homogenous in terms of symptom-generating mechanism than the entire group of FD.²⁰ Recently, duodenal pathology, such as eosinophilic infiltration, has been suggested as an important mechanism for FD.^{14,21} Accordingly, it is essential to determine if the conventional symptom-based subtypes could also reflect the status of the duodenal pathology. The eosinophils and mast cell counts in the duodenum and stomach were examined according to symptom-based subtypes of FD, and no differences were found among the subtypes. If the duodenal pathology is an important symptom-generating mechanism of FD, it should be utilized in the treatment strategies. Nevertheless, these findings show that the conventional symptom-based subtypes used for an FD treatment strategy do not reflect the duodenal pathology. Although more confirmative studies are required, the symptom-based subtypes will need to be modified.

Duodenal eosinophilic infiltration may induce gastroduodenal dysmotility via eosinophil-induced T-cell activation and subsequent release of leukotrienes.²²⁻²⁴ Indeed, several studies have shown increased duodenal eosinophil counts in patients with FD compared to healthy controls.^{12,13,25} On the other hand, some studies reported no significant association

between duodenal eosinophilia and FD. Therefore, the presence of duodenal eosinophilic degranulation may be more important.^{25,26} In an Australian study, increased duodenal mucosal eosinophil counts were not associated with FD but with postprandial symptoms (postprandial fullness and early satiety).¹² By contrast, the present study found no association between the duodenal eosinophilic count and symptom-based subtypes of FD. Mast cell infiltration could be involved in the symptom-generating mechanism of FD.^{13,27} Duodenal mast cell infiltration may be linked to IBS.²⁸ In the present study, an increased tendency of the mast cell count was observed in the overlap group. A possible explanation is that the overlapping group may share IBS characteristics.

This study had some limitations. This study was retrospective. Thus, the results are limited in terms of generalization. Eosinophil degranulation was not investigated. Activated eosinophils may be more important in the FD pathophysiology. Nevertheless, this study investigated the differences in eosinophil and mast cell infiltration in patients with FD according to symptom-based subtypes. In addition, the present study has a small sample size and no healthy control data. Although the differences in eosinophil and mast cell infiltration in the duodenum and stomach were examined according to symptom-based subtypes of FD, comparable data may be obtained using a control group. Subjects with a *H. pylori* infection by staining may be excluded to reduce confounding effects. Nevertheless, it would be better to consider the past eradication history together. The small sample size lowers the power of the results and limits detailed analysis, but the negative observations could help design future large prospective studies.

In conclusion, the duodenal eosinophil and mast cell counts were similar according to the symptom-based subtypes of FD. This suggests that the currently used subtyping of FD does not reflect the duodenal eosinophil and mast cell infiltration. Thus, a new subtyping of FD must include novel emerging pathophysiology.

REFERENCES

1. Talley NJ, Stanghellini V, Heading RC, Koch KL, Malagelada JR, Tytgat GN. Functional gastroduodenal disorders. Gut 1999;45 Suppl 2(Suppl 2):II37-II42.
2. Tack J, Bisschops R, Sarnelli G. Pathophysiology and treatment of functional dyspepsia. Gastroenterology 2004;127:1239-1255.
3. Doran S, Jones KL, Andrews JM, Horowitz M. Effects of meal vol-

- ume and posture on gastric emptying of solids and appetite. *Am J Physiol* 1998;275:R1712-R1718.
4. Mundt MW, Hausken T, Samsom M. Effect of intragastric barostat bag on proximal and distal gastric accommodation in response to liquid meal. *Am J Physiol Gastrointest Liver Physiol* 2002;283:G681-G686.
 5. Karamanolis G, Caenepeel P, Arts J, Tack J. Association of the predominant symptom with clinical characteristics and pathophysiological mechanisms in functional dyspepsia. *Gastroenterology* 2006;130:296-303.
 6. Bouras EP, Delgado-Aros S, Camilleri M, et al. SPECT imaging of the stomach: comparison with barostat, and effects of sex, age, body mass index, and fundoplication. Single photon emission computed tomography. *Gut* 2002;51:781-786.
 7. Stanghellini V, Chan FK, Hasler WL, et al. Gastroduodenal disorders. *Gastroenterology* 2016;150:1380-1392.
 8. Enck P, Azpiroz F, Boeckxstaens G, et al. Functional dyspepsia. *Nat Rev Dis Primers* 2017;3:17081.
 9. Miwa H, Kusano M, Arisawa T, et al. Evidence-based clinical practice guidelines for functional dyspepsia. *J Gastroenterol* 2015;50:125-139.
 10. Oh JH, Kwon JG, Jung HK, et al. Clinical practice guidelines for functional dyspepsia in Korea. *J Neurogastroenterol Motil* 2020;26:29-50.
 11. Walker MM, Salehian SS, Murray CE, et al. Implications of eosinophilia in the normal duodenal biopsy - an association with allergy and functional dyspepsia. *Aliment Pharmacol Ther* 2010;31:1229-1236.
 12. Walker MM, Aggarwal KR, Shim LS, et al. Duodenal eosinophilia and early satiety in functional dyspepsia: confirmation of a positive association in an Australian cohort. *J Gastroenterol Hepatol* 2014;29:474-479.
 13. Wang X, Li X, Ge W, et al. Quantitative evaluation of duodenal eosinophils and mast cells in adult patients with functional dyspepsia. *Ann Diagn Pathol* 2015;19:50-56.
 14. Wauters L, Talley NJ, Walker MM, Tack J, Vanuytsel T. Novel concepts in the pathophysiology and treatment of functional dyspepsia. *Gut* 2020;69:591-600.
 15. Talley NJ, Tack J, Ptak T, Gupta R, Giguère M. Itopride in functional dyspepsia: results of two phase III multicentre, randomised, double-blind, placebo-controlled trials. *Gut* 2008;57:740-746.
 16. Hallerbäck BI, Bommelaer G, Bredberg E, et al. Dose finding study of mosapride in functional dyspepsia: a placebo-controlled, randomized study. *Aliment Pharmacol Ther* 2002;16:959-967.
 17. Tack J, Van Den Elzen B, Tytgat G, et al. A placebo-controlled trial of the 5-HT_{1A} agonist R-137696 on symptoms, visceral hypersensitivity and on impaired accommodation in functional dyspepsia. *Neurogastroenterol Motil* 2009;21:619-e24.
 18. Hill LD, Kozarek RA, Kraemer SJ, et al. The gastroesophageal flap valve: in vitro and in vivo observations. *Gastrointest Endosc* 1996;44:541-547.
 19. Kimura K, Satoh K, Ido K, Taniguchi Y, Takimoto T, Takemoto T. Gastritis in the Japanese stomach. *Scand J Gastroenterol Suppl* 1996;214:17-23.
 20. Lee KJ. The usefulness of symptom-based subtypes of functional dyspepsia for predicting underlying pathophysiologic mechanisms and choosing appropriate therapeutic agents. *J Neurogastroenterol Motil* 2021;27:326-336.
 21. Moshiree B, Talley NJ. Functional dyspepsia: a critical appraisal of the European consensus from a global perspective. *Neurogastroenterol Motil* 2021;33:e14216.
 22. Rothenberg ME, Cohen MB. An eosinophil hypothesis for functional dyspepsia. *Clin Gastroenterol Hepatol* 2007;5:1147-1148.
 23. Gargala G, Lecleire S, François A, et al. Duodenal intraepithelial T lymphocytes in patients with functional dyspepsia. *World J Gastroenterol* 2007;13:2333-2338.
 24. Liebrechts T, Adam B, Bredack C, et al. Small bowel homing T cells are associated with symptoms and delayed gastric emptying in functional dyspepsia. *Am J Gastroenterol* 2011;106:1089-1098.
 25. Lee MJ, Jung HK, Lee KE, Mun YC, Park S. Degranulated eosinophils contain more fine nerve fibers in the duodenal mucosa of patients with functional dyspepsia. *J Neurogastroenterol Motil* 2019;25:212-221.
 26. Järbrink-Sehgal ME, Sparkman J, Damron A, et al. Functional dyspepsia and duodenal eosinophil count and degranulation: a multiethnic US veteran cohort study. *Dig Dis Sci* 2021;66:3482-3489.
 27. Yuan HP, Li XP, Yang WR, Li FK, Li YQ. Inducible nitric oxide synthase in the duodenal mucosa is associated with mast cell degranulation in patients with functional dyspepsia. *Ann Clin Lab Sci* 2015;45:522-527.
 28. Walker MM, Talley NJ, Prabhakar M, et al. Duodenal mastocytosis, eosinophilia and intraepithelial lymphocytosis as possible disease markers in the irritable bowel syndrome and functional dyspepsia. *Aliment Pharmacol Ther* 2009;29:765-773.